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Impairment of the Reproductive System and Mating of Male and Female Black Rats Treated with Lambda-Cyhalothrin Bait and Its Field Application

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The impact of lambda-cyhalothrin (LCT) bait on the reproductive system of adult male and female black rats (Rattus rattus) as well as the mating ability and its field efficiency were evaluated in the current study. Testis, epididymis and ovary were weighed, and some oxidative stress and antioxidant parameters of treated and untreated rats were assessed after feeding on LCT (0.024 %) for four days. Moreover, male and female hormones were measured, and sperm count, motility and vitality percent were recorded. Sperm abnormalities and histological changes of the testis and ovary were examined. Pregnancy percent and fetus numbers were recorded. Field application of LCT bait (0.032 %) was evaluated in two successive years. The results revealed that body, testes, epididymis and ovary weights decreased in treated rats compared with the untreated. Testicular and ovarian malondialdehyde (MDA) activity was increased while glutathione content and superoxide dismutase level was decreased compared with untreated rats. Regarding the hormones, testosterone, LH, FSH, and progesterone, estrogen levels were depleted in treated rats. Additionally, LCT bait caused a reduction in sperm count, motility and vitality percent compared with the control. LCT bait motivated deformations in sperm forms including detached head, hookless head, bent neck and tail as well as coiled tail. In addition, tissue impairments in the testis and ovary were observed. Concerning mating ability, the compound caused decreasing in the pregnancy percentage, and fetus number. Moreover, LCT bait induced a remarkable reduction in the rat population recording 70.36 and 68.72 % in the two study years, respectively under field conditions. Finally, it was concluded that LCT bait has an effective impact on the reproductive systems of both male and female rats and reducing pregnancy rates as well as population reduction in field application. So, we suggest involving LCT bait in the integrated control program for rodents.

INTRODUCTION

Rodents cause massive economic losses worldwide through consumption, damaging and contaminating crops in the field as well as during storage. In addition, rodents are carriers of various diseases to animals and humans (John, 2014). Lambda-cyhalothrin (LCT) is a synthetic pyrethroid insecticide. It is used for the control of various insects in agriculture (Ghramh *et al.*, 2022). Moreover, subsequent studies have revealed that LCT is toxic to mammals (Kandil *et al.*, 2022a; Settar *et al.*, 2023). Mate *et al.* (2010) recorded that the oral LD₅₀ of LCT is 79 and 56 mg/kg for male and female rats, respectively. LCT can block the closing of neuronal sodium ion channels and change the normal function of nerves

in insects and mammals causing paralysis or death (Tomar et al., 2015; Lopez-Torres et al., 2022). The toxicity degree depends on its dose, method and duration of exposure. LCT is known as a strong agent for biochemical distortion, nervous disruption and oxidative damage (Ezenwosu et al., 2021; Alrawe and ALzibaidy, 2022). Kandil et al. (2022a) reported that LCT bait (0.032 %) has rodenticidal efficacy under laboratory and field conditions against black rats (Rattus rattus). Oral administration of LCT caused a reduction in the body weight of albino rats (Al-Amoudi, 2018). Oxidative stress is considered one of the major mechanisms by which pesticides alter reproduction (Sule et al., 2022). The oxidative stress process occurs when the reactive oxygen species (ROS) generation exceeds the antioxidant scavenging capacity in organisms (Juan et al., 2021). Administration of LCT at high doses (4.16 and 8.32 mg/kg b.wt) has potentially adverse effects on the female rabbit via decreased serum progesterone, decreased number of implantation sites, placenta and litter size, live fetus as well as an increased number of dead fetuses and total resorptions (Adam et al., 2019). Reported studies revealed male reproductive toxicity in the form of decreased semen quality as well as decreased testicular and epididymal weights in rabbits (Yousef, 2010). Additionally, its exposure causes sperm abnormalities, reduction in sperm count and motility in rats as well as blocked spermatogenesis and damaged seminiferous tubules in mice exposed to LCT (Al-Sarar et al., 2014). It was reported that pyrethroids could affect ovulation causing atresia of follicles, decreasing follicular cells, oocytes and corpora lutea as well as inducing vesicular atrophy of the endometrial glands of mammals (Kotil and Yön, 2015; Marettova et al., 2017). Previous studies have indicated that several pyrethroids induce toxic impacts on the reproduction of mammals (Marettova et al., 2017; Abdel- Maksoud et al., 2018; Zhang et al., 2021; Zhong et al., 2021). Pyrethroids and their metabolites showed toxic effects on the endocrine system causing disturbances in the female hormones of mammals (Fei et al., 2010; Liu et al., 2011; Jurewicz et al., 2020). Reproduction is a vital function that permits species continuity. It enhances productivity and eternizes animal species and its disturbance or dysfunction causes negative effects on animal productivity (Fernandez-Novo et al., 2020; Abd El-Rheem et al., 2022). This research aimed to assess the effect of LCT bait on the reproductive system of male and female rats, follow up the mating ability and pregnancy to find out its effect on the production of new individuals as well as the extent of its field application during two successive years and the possibility of using it as a rodenticide.

MATERIALS AND METHODS

Tested Compound:

Common Name: Lambda-cyhalothrin, oral LD_{50} for rats is 79 mg/kg (World Health Organization, 1990 and Mate *et al.*, 2010)

Trade name: Dolf-X 5% EC was purchased from Star Chem Chemical Manufacturing Co., 6th of October City, Giza, Egypt. It was used as bait mixed with crushed maize and used sunflower oil at a level of 0.024 % as a sublethal concentration for biochemical tests and another concentration of 0.032 % for field application according to Kandil *et al.* (2022a). **Laboratory Experiments:**

Experimental Animals: Using rat traps $(30 \times 15 \times 20 \text{ cm})$, black rats, *Rattus rattus*, were caught from fields in Beba district, Beni-suef Governorate, Egypt. Then rats were transported to the Laboratory of Harmful Animals Researches, Sids Agriculture Research Station, ARC. Animals were put individually in cages $(50 \times 30 \times 30 \text{ cm})$. Animals were offered crushed maize and water at 20-25°C and 12 hours of daily light/dark cycles for 15 days before the start of the experiment. Only mature healthy rats (males and females) were selected for the following experiments.

Experimental Design: One hundred and thirty adult healthy black rats, Rattus rattus were

divided as follows:

a. Groups for Biochemical Studies:

Forty rats were weighed 170-190 g/kg b.wt., and divided into 2 groups (10 males & 10 females of each). The first group was fed on LCT- bait (0.024 %) for four successive days according to (Kandil *et al.*, 2022a). The second group was fed crushed maize without treatment as a control. These groups were used to determine some oxidative stress and antioxidant enzymes, hormones as well as sperm count, morphology, motility and vitality percent. In addition to that, histopathological changes were investigated.

b. Groups for Mating Ability Studies:

Ninety rats were used to assess mating ability and follow pregnancy, animals were divided into sixty rats (20 males and 40 females) treated with LCT (bait 0.024%) and the other thirty rats (10 males and 20 females) as untreated. After 4 days of treatment, killed animals were excluded and the surviving animals were divided into four groups (each of 5 males and 10 females). Group 1: untreated males mated with untreated females as a control. Group 2: untreated males mated with treated females. Group 3: treated males mated with untreated females. Females of each group were daily checked for pregnancy. After 20 days, the females were dissected, and the numbers of implantation and resorption sites as well as embryo numbers were recorded.

Samples Preparation: After treatment; the surviving rats were weighed and samples were collected.

- **Blood Samples:** Blood samples were collected from a cervical vein, and centrifuged for 10 min at 3000 rpm. The clear supernatant serum was kept in a deep freezer at -20°C until use (Henry *et al.*, 1979).
- **Tissue Preparations:** Right testis and right ovary of rats were separated, and homogenized with 0.9% NaCl (each 0.5 g tissue in 5 ml 0.9% NaCl). Then, it was centrifuged at 3000 rpm for 15 minutes. The supernatants were kept in a deep freezer at -20°C till use.
- Semen Collection: The right cauda epididymids was extirpated and cut with a sterilized scissor in Petri- dish where the spermatozoa were dispersed in 2 ml pre-warmed physiological saline solution at 37C° (Kempinas *et al.*, 1988). The same processes were done with the untreated group.

1. Biochemical Analysis:

- Oxidative Stress, Antioxidants and Hormonal Assay: Glutathione (GSH) content, superoxide dismutases (SOD) and Malondialdehyde (MDA) levels (in testis and ovary) were assessed using a reagent kit purchased from Biodiagnostic Company (Egypt) according to (Beulter *et al.*, 1963), (Nishikimi *et al.*, 1972) and (Ohkawa *et al.*, 1979) methods, respectively. Serum testosterone in male rats, luteinizing hormone (LH), follicle-stimulating hormone (FSH) in both (males & females) and progesterone levels in females were estimated by rat enzyme-linked immunosorbent assay (ELISA) commercial kits purchased from Cusabio Biotech CO., Wuhan, China, according to the manufacturer's protocol. Estrogen level was estimated by rats enzyme-linked immunosorbent assay (ELISA) commercial kits purchased from Bioassay Technology Laboratory (Rat Estrogen Elisa kit), China, according to the manufacturer's protocol.
- Sperm Count, Motility Percent and Morphology: Sperm count, sperm motility and vitality percentage were analyzed using assisted sperm analysis (CASA) of rat epididymal spermatozoa (Amann and Waberski, 2014). The sperm motility was expressed in a percentage of motile sperm while sperm count was expressed as million sperm cells per ml of suspension. Morphological abnormalities of sperms were detected using the method of (Filler, 1993; Ardıç *et al.*, 2021; Ahmed *et al.*, 2023).

2. Histological Investigations: According to the procedure of (Banchroft *et al.*, 1996), the left testis and left ovary were quickly removed after sacrifice, decapitation, and dissection and then, they were perfused with saline solution. After that, the specimens were fixed in 10% neutral buffered formalin for 24 hours and transferred to the Pathology Laboratory of Animal Health Research Institute, Giza, Egypt for histological investigations. The

histological hematoxylin and eosin (H&E) stained testis and ovary sections were examined to detect the histopathological lesions.

Field Application: LCT bait (0.032 %) was evaluated under the field conditions during two seasons (the year 2020 and 2021) at Quftan Village, Beba district, Beni-Suef Governorate, Egypt according to the method described by Dubock (1984). This bait concentration (0.032 %) was chosen according to the previous study (Kandil et al., 2022a). The trials were conducted during November after the summer crops harvest in maize fields as an infested area. Three plots (each of one fedden) were used for treatment and the other three plots were left as a check control area. The population density of the rodent species was estimated pre and post-treatment using the crushed maize consumption method. For pre-treatment, crushed maize (in small plastic sacks 50 gm of each) was put inside bait stations and distributed in every plot for 5 successive days. The average consumed amount of food was calculated for only the last two days. After that, 50 grams of the LCT bait was offered to rats inside the bait stations and the consumed quantity of the bait was replaced every week for three weeks, with a recording of the consumption. After that, the bait stations were left empty for one week. Then untreated crushed maize was placed and the consumed amount was calculated as above in the pretreatment period. The reduction in rodent population was estimated as follows:

Population reduction % = $\frac{\text{Pre} - \text{treatment consumption } (g) - \text{Post} - \text{treatment consumption } (g)}{(\text{Pre} - \text{treatment consumption}(g))} x100$

Statistical Analysis: The experimental design was completely randomized with different replicate. The obtained results were statically analyzed by one-way analysis of variance (ANOVA) and least significant difference (LSD) at (P < 0.05) using the COSTAT program (Glenn, 2005)

RESULTS

Effect of LCT Bait (0.024 %) on Body, Testes, Epididymis and Ovary Weights of Black Rats:

As depicted in Table (1), after male and female rats fed on LCT bait (0.024%) the results showed significant decreases (p < 0.05) in the body of male and female, testes and epididymis weights with -5.84, -5.96 %, -20.72 and -5.88 % difference, respectively compared with controls. While treatment induced an insignificant decrease (p > 0.05) in the ovary weight of rats with a -1.35 % difference compared with untreated rats.

		2			
Organ weight (g)		Mean ± SE	of weight (g)	Difference	LCD
		Control	Treatment	%	LSD
Body	Male	198.6±3.33ª	187.0±3.11 ^b	-5.84	10.53
	Female	198.0±3.79ª	186.2±2.65 ^b	-5.96	10.68
Testes		2.22±0.09ª	1.76±0.07 ^b	-20.72	0.253
Epididymis		0.34±0.01ª	0.32±0.01 ^b	-5.88	0.019
Ovary		0.148±0.01ª	0.146±0.01ª	-1.35	

Table 1: Effect of lambda-cyhalothrin bait (0.024 %) on body, testes, epididymis and ovary weights of black rats after four days of treatment.

Values are expressed as means± standard error

^{ab} values with different letters are significantly different at (p< 0.05). LSD: Least Significant Difference

Biochemical Changes:

Effects On Some Oxidative Stress and Antioxidant Parameters:

LCT bait (0.024%) effects on some oxidative stress (MDA) and antioxidants (GSH and SOD) of male and female rats are shown in Table (2). Testicular and ovarian MDA level was increased by 59.95 and 48.85 % difference, respectively in treated rats compared

to untreated rats. Moreover, GSH and SOD levels recorded a -20.83, -19.33% difference, respectively in testicular tissues and a -17.71%, and -21.00 % difference in ovarian tissues compared with controls.

Table 2: Lambda-cyhalothrin bait (0.024%) effect on some oxidative stress and antioxidant parameters level of black rats.

Parameter	Organ	Control	Treatment	Difference %	LSD
MDA (nm/g tissue)	Testis	7.64±0.31 ^b	12.22 ± 0.64^{a}	59.95	1.65
	Ovary	4.77±0.17 ^b	7.10±0.12ª	48.85	0.478
GSH (mg/g tissue)	Testis	28.8±1.02ª	22.8±1.49 ^b	-20.83	4.18
	Ovary	19.2±1.24ª	15.8±0.73 ^b	-17.71	3.33
SOD (U/g tissue)	Testis	23.8±1.35ª	19.2±0.49 ^b	-19.33	3.33
	Ovary	20.0±1.30ª	15.8±1.11 ^b	-21.00	3.95

Values are expressed as means± standard error

^{ab} values with different letters are significantly different at (p < 0.05). LSD: Least Significant Difference.

Effects of LCT Bait (0.024%) on Sex Hormones of Male and Female Black Rats:

As presented in Table (3), feeding with LCT bait (0.024%) motivated a significant reduction in testosterone, LH and FSH levels of male rats with -10.06, -16.13 and -40.00 % difference, respectively compared with control. While progesterone, estrogen, LH and FSH levels of female rats recorded -33.33, -25.48, -55.81 and -64.80 % difference, respectively compared with controls.

Table 3: Lambda-cyhalothrin bai	t (0.024%) effect	on male and fema	le sex hormones of
black rats.			

Hormone	Sex	Control	Treatment	Difference %	LSD
Testosterone (ng/ml)	Male	1.59±0.01ª	1.43±0.02b	-10.06	0.071
Progesterone (ng/ml)	Female	1.26±0.03ª	0.84±0.02 ^b	-33.33	0.093
Estrogen(ng/L)		4.20±0.26ª	3.13±0.09 ^b	-25.48	0.746
LH (mIU/ml)	Male	0.93±0.03ª	0.78±0.02 ^b	-16.13	0.093
	Female	1.72±0.05ª	0.76±0.03 ^b	-55.81	0.172
FSH (mIU/ml)	Male	0.55±0.02ª	0.33±0.02b	-40.00	0.077
	Female	1.25±0.05ª	0.44±0.04 ^b	-64.80	0.173

Values are expressed as means± standard error

^{ab} values with different letters are significantly different at (p < 0.05). LSD: Least Significant Difference.

Effect of LCT Bait (0.024%) on Count, Vitality and Motility % of Sperm of Male Black Rats:

The impact of LCT bait on sperm count, vitality and motility percent of sperm of rats are shown in Table (4). Sperm count significantly decreased in treated male rats (26.23 $\times 10^6$ spermatozoa/ml) compared with control rats (81.23 $\times 10^6$ spermatozoa/ml). In addition to that, sperm vitality and motility percent decreased in a significant manner to 21.87 % and 24.33 %, respectively, compared with 76.86% and 87.36 % of controls.

Group	Group Sperm count (×10 ⁶ spermatozoa/ml)		Motility (%)	
Control	81.23±1.36ª	76.86±0.95ª	87.36±5.64ª	
Treatment	26.23±1.78 ^b	21.87±0.95 ^b	24.33±1.77b	
LSD	6.21	3.73	16.39	

Table 4: Effect of lambda-cyhalothrin bait (0.024%) on count, vitality and motility% ofsperm of male black rats.

Values are expressed as means \pm standard error

 ab values in column with different letters are significantly different at (p<0.05).

LSD: Least Significant Difference

Effect of LCT Bait (0.024%) on Sperm Morphology:

LCT bait (0.024%) caused various deformations in the head, neck and tail of rat sperms. These malformations include a hookless head (Figs. 2& 4) as well a detached head and bent neck (Figs. 3& 4), cytoplasmic droplets (Fig. 4) as well a coiled tail (Fig. 5) and a bent tail (Fig. 6). Comparison with the normal form of sperm with normal head, neck and tail were observed in untreated rats (Fig. 1).

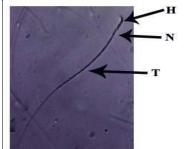


Fig. 1: Photomicrograph of sperm of untreated showing normal morphological sperm of untreated rats showing normal head (H), neck (N) and tail (T) (Nigrosin-Eosin stain, 400 x).

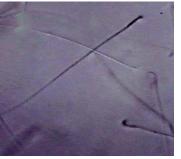


Fig. 2: Photomicrograph of sperm of LCT treated rats showing hookless head (Nigrosin-Eosin stain, 400 x)

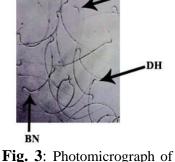


Fig. 3: Photomicrograph of sperms of LCT treated rats showing bent neck (BN) and detached head (DH). (Nigrosin-Eosin stain, 400 x).

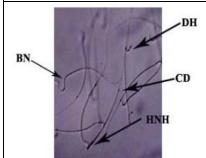


Fig. 4: Photomicrograph of sperms of LCT treated rats showing bent neck (BN) and detached head (DH), cytoplasmic droplets (CD) and hookless head (HNH) (Nigrosin-Eosin stain, 400 x).



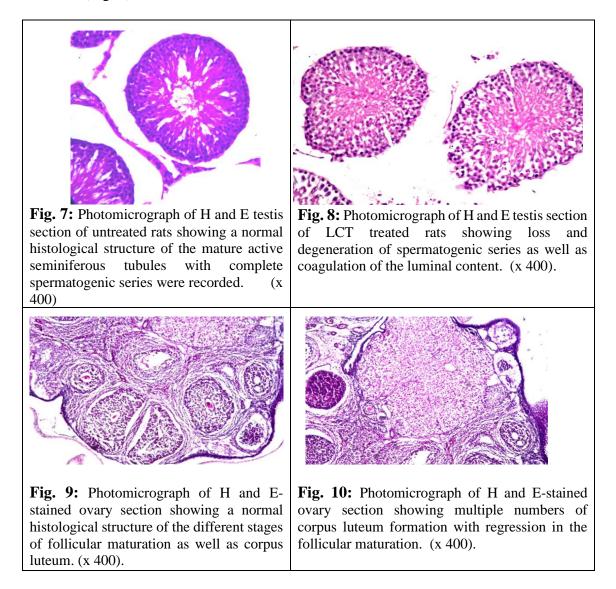
Fig. 5: Photomicrograph of sperms of LCT treated rats showing coiled tail. (Nigrosin-Eosin stain, 400 x)



Fig. 6: Photomicrograph of sperm of LCT treated rats showing bent tail (Nigrosin-Eosin stain, 400 x).

Histopathological Studies:

Sections of the testis showed a systematic histological structure. Normal spermatogonial cells, spermatids and sperms were depicted in untreated male rats (Fig. 7). On the other hand, LCT bait (0.024%) caused histopathological changes including degeneration of the seminiferous tubules and loss of spermatogenic series with coagulation of the luminal content (Fig. 8). Regarding the impact of LCT bait on the ovary of rats, it showed multiple numbers of corpus luteum formation with regression in the follicular maturation (Fig. 10) compared to the ovary of untreated rats showed the normal histological structure of the different stages of follicular maturation as well as corpus luteum were observed (Fig. 9).



Effect of LCT (0.024%) on Mating Ability and Pregnancy:

Table (5) and Figures (11, 12, 13) show the effect of LCT bait (0.024 %) on the pregnancy percentage and fetus. The results cleared that LCT bait caused a reduction in pregnancy percent compared with untreated rats whereas, G2 (Mating of untreated males with treated females), G3 (treated male with untreated females), and G4 (treated males with treated females) induced 40, 20, and 10 % pregnancy comparing with 80 % of controls G1(untreated males with untreated female). Moreover, it caused a reduction in the number of the fetus in treated rats compared with untreated pregnant rats.

	of fetuses after 20 days gestation of futs.							
Group	Pregnancy %	No of implantation	No of resorption sites/	Total number of				
		sites	pregnant rat	fetuses				
G1	80	7.2±1.20 ª	0	72				
G2	40	2.8±1.14 b	0	28				
G3	20	1.4±0.93 b	0	14				
G4	10	0.7±0.70 b	0	7				
LSD		2.93						

Table 5: Effect of lambda-cyhalothrin bait (0.024 %) on pregnancy percent and the numberof fetuses after 20 days gestation of rats.

G1: untreated males mated with untreated females as a control. G2: untreated males mated with treated females. G3: treated males mated with untreated females. G4: treated males mated with treated females. Values are expressed as means \pm standard error ^{ab} values in column with different letters are significantly different at (p< 0.05). LSD: Least Significant Difference.



Field Application:

The efficiency of LCT bait (0.032%) was evaluated against black rats as shown in Table (6). Regarding the pre-treatment period in the first and second year, the crushed maize average consumption before treatment was1383.33 and 1183.33 g, respectively, while in post-treatment, the consumption was 410.00 and 370.00 g, and consumption of treated bait was 750.00 and 706.00 g, respectively. Our data detected that LCT bait achieved a 70.36 and 68.72 % reduction in the rat population in the two years, respectively. It was recorded significant difference at (p< 0.05) between average rat consumptions during the experiment period in each year.

Average consumption of bait (g) Mean ± SE					Population reduction	LSD
Year	Pre-Treatment	Treatment	Post- Treatment	Control	%	LSD
2020	1383.33±27.32ªA	750.00±50.39 ^b	410.00±15.29°	1440.00±32.18 ^{aA}	70.366	109.92
2021	1183.33±8.83 ^{aB}	706.00±31.84 ^b	370.00±17.34°	1190.00±36.10 ^{aB}	69.70	01 55
LSD	79.59			111.42	68.72	84.55

Table (6): Efficiency of lambda-cyhalothrin bait (0.0.032 %) against black rats under field conditions.

Values are expressed as means \pm standard error ^{abc} values with different letters are significantly different at (p< 0.05). ^{A, B} values in a column with different letters are significantly different at (p< 0.05). LSD: Least Significant Difference

DISCUSSION

In the current study, LCT bait caused a noticeable decrease in body weights of male and female black rats and this might be related to LCT bait toxicity that decreased food

intake and absorption via the gastrointestinal tract and also affected food conversion. These results are in parallel with those of (Al Malahi et al., 2022), who revealed that feeding with different concentrations of LCT induced a reduction in body weight of male and female mice and this returned to LCT decreased the dietary intake. Testicular and epididymis weights are an important index of reproductive health. The present study detected that testicular and epididymis weights decreased in treated rats and this decrease may be returned to the reduction in the number of germ cells, induction of oxidative stress and spermatogenesis inhibition as well as a reduction in testosterone level. These data are in accordance with Zhang et al. (2021) who concluded that pyrethroid exposure caused a reduction in testicular and epididymis weights of male mice. The reduction in ovarian weight of LCT-treated rats observed in this study may be due to the reduction in the number of ovarian germ cells. In this research, LCT bait feeding increased MDA levels in the testis, revealing an increase in tissue oxidative stress. This may be a result of an accumulation of LCT in testicular and ovarian tissues causing membrane degeneration and immoderate formation of free radicals. Too many free radicals damage the membranes and the antioxidant defense of the tissue and this accelerates oxidative stress, which appeared in form of increased MDA levels on LCT feeding. This illustration is supported by the degeneration of spermatogenic series in histological studies. These results are in parallel to those of Ben Abdalla et al. (2013) who revealed that LCT exposure caused a decrease in testicular MDA level as a result of free radicals induction and elevation of lipid peroxidation. It was reported previously that pesticides promote an increase in lipid peroxidation levels (Saad-Hussein et al., 2019; Sule et al., 2022). Concerning GSH is a powerful antioxidant that is serious for cellular protection preventing damage to important cellular components caused by free radicals and peroxides (Matuz-Mares et al., 2021). In our study, testicular and ovarian GSH reduction was observed after LCT bait feeding. This reduction may be related to enhanced usage of GSH for detoxification of LCT bait-induced free radicals. Our results are in accordance with those of Ghosh et al. (2016) who mentioned that LCT causes depletion in GSH content due to its usage to challenge oxidative stress. Superoxide dismutase (SOD) is regarded as the first line of defense against the harmful effects of reactive oxygen species (ROS) in the cell by catalyzing the dismutation of superoxide radicals to produce H₂O₂. Suppressions in testicular and ovarian SOD activities were observed in LCT-treated rats and this may be a result of SOD exhausting to combat the oxidative stress induced by LCT bait. So depletion of antioxidants in our study suggested the harmful impact of LCT bait in tested rats. Previous studies proved that several antioxidants can combat the negative impacts of various pesticides (Abdel-Tawwab et al., 2021; Abdul Ghani and Naser, 2022). The oxidative damage that occurred in the present study after the feeding of LCT bait is supported by the histological changes in the testis and ovary of black rats. These data are in parallel with those of (Al-Sarar et al., 2014; Al Malahi et al., 2022). In our work, LCT bait caused degeneration of the seminiferous tubules and loss of spermatogenic series with coagulation of the luminal content of male rats. These impairments may be a result of the impact of ROS produced by LCT and the reduction of testosterone. On the other hand, multiple numbers of corpus luteum formation with regression in the follicular maturation in LCT-treated rats. It was also revealed that LCT causes cytotoxic and genotoxic impacts (Shen et al., 2020).

Normal reproductive function is under the effect of sex hormones. In the present work, LCT bait motivated a remarkable lowering in testosterone, LH, FSH, progesterone, and estrogen level and this may be a return to damage of cells as a result of induction of oxidative stress destroying the membranes of cells. These data are supported by the histological studies which revealed degeneration of the testicular and ovarian cells. These results are in parallel with Abdel- Maksoud *et al.* (2018), Ghosh *et al.* (2018) and Abd El-Hameed and Mahmoud (2020) who indicated that pyrethroids enhance suppression in male

and female hormones. Testosterone is important to maintain both of structure and function of the male accessory sex gland. Reduction of testosterone impairs spermatogenesis (Grande *et al.*, 2022) where its biosynthesis of testosterone is mainly located in testicular Leydig cells after being stimulated by LH, which is secreted from the pituitary gland as a result of the pulsatile releasing of gonadotropin-releasing hormone (GnRH) from the hypothalamus (Smith and Walker, 2014). In addition, FSH secreted by the pituitary acts on the sertoli cells of the testes to facilitate spermatogenesis. Various hormones generated by the hypothalamic-pituitary complex and also the sex organs have essential roles in the control of folliculogenesis, oogenesis and ovulation (Jones and Shikanov, 2019).

In addition to that, LCT bait caused a significant decrease in sperm count, vitality and motility percent as well as sperm deformations and this may be due to the oxidative damage of LCT bait destroying the testicular cells affecting the spermatozoa production as well as the coagulation in the seminiferous tubules may lead to sperms death and reduction in testis and epididymal weights. The epididymis plays an essential role in sperm maturation and storage in mammals. During epididymal transit, sperm metabolism elevates but is accompanied by exposure to oxidative stress (Dacheux and Dacheux, 2014). These data are in accordance with Al Malahi *et al.* (2022) who reported that LCT caused damage and change in the sperm morphology of albino mice as a result of the destruction of the testicular cells. So in this study, accumulation of LCT bait in the testis induces oxidative stress accelerating the death of spermatogenic cells as well as may associate with sperm abnormalities. Concerning the mating ability, the suppression in the mating ability of treated rats may return to hormonal disturbance, tissue impairments of the testis and ovary as well as a decrease in sperm count, vitality and motility after the feeding of LCT bait and induction of oxidative stress.

Regarding field application, LCT bait achieved a 70.36 and 68.72 % reduction in the rat population in two successive years, respectively. This may be due to the toxic impact of LCT bait causing the destruction of cells and death. These results are in accordance with our previous study (Kandil *et al.*, 2022a) as LCT bait achieved a 71.34% reduction in the rat population in crop stores. This reveals the efficiency of the LCT bait under field conditions. Kandil *et al.* (2022b) stated that spirotetramat achieved an 80 % reduction in the rat population under field conditions.

Conclusion

LCT has a strong effect on the reproductive system of male and female rats. It caused the destruction of testis and ovary tissues and serious disorders of antioxidant enzymes and hormones responsible for the regulation of the reproduction process. Moreover, it decreased the incidence of pregnancy and the number of fetuses. In addition to that, LCT bait achieved efficiency against rats under field conditions whereas, it achieved 70.36 and 68.72 % reduction percent during two years, respectively in this study and gave a 71.3% reduction in the population of rats in the previous study, so it investigated that it can achieve average 70% population reduction when it includes in the integrated rodent control in Egypt.

Conflicts of Interest:

All the authors have no competing conflicts of interest.

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