Effect of Formaldehyde on The Structure of The Lung and Heart and The Possible Protective Effect of Omega- 3 (Histological and immune-histochemical study)

Salwa M. Ouies and Abeer F. Abd El-Naeem
Human anatomy & Embryology Department, Faculty of Medicine, Sohag University
Email: salwaouies@yahoo.com - abeerfareed_2013@yahoo.com

ABSTRACT
Background: Formaldehyde (FA) is a common indoor and outdoor pollutant found in many products. The toxicity of formaldehyde is of concern to all who work closely with it such as embalmers, anatomists, histology technicians, and medical students. Omega-3 in fish oil is one of the most important polyunsaturated fatty acids (PUFA) that have anti-inflammatory and antioxidant activity. Materials and methods: Thirty adult male albino rats were divided into three equal groups. Group I was a control group. Group II rats were exposed to formaldehyde inhalation for 4 weeks. Group III were exposed to formaldehyde inhalation and were administered orally with a 300mg/kg/day Omega-3 for 4 weeks. After 24 hr of the last dose, the animals were dissected. Hearts and lungs were processed for examination after Haematoxylin and eosin, Masson trichome and immunohistochemical stains. Results: The rats treated with formaldehyde inhalation showed significant changes in the normal architecture of both lung and heart. The lung showed congestion and mass appeared on one lobe and increase in the area percent of collagen fibers. Immune-expression of tumor necrosis factor-alpha (TNF-α), and caspase-3 were increased in the lung and heart compared to the control. Omega-3 fatty acids can ameliorate the pathological changes, decrease the fibrosis and the immune-expression in both lung and heart. Conclusion: Formaldehyde was associated with many histopathological changes in both lung and heart and the Omega-3 can ameliorate these changes.

INTRODUCTION
Formaldehyde is an organic carbon compound frequently used in occupational environments (hospitals, textiles, paper, resins, and wood composites), house indoor environments (insulating materials, fabrics, chipboard, and cooking emissions), and also in anatomy, pathology, and histology laboratories. (Kim et al., 2002; Nakazawa et al., 2005; Leal et al., 2018).

Formaldehyde inhalation inflicts various harms on many systems, not include the respiratory system only but also exerted on a variety of organs of living bodies (Zhou et al., 2006). Exposure of the medical students and instructors to formaldehyde during the gross anatomy laboratory may cause an acute effect on the respiratory system and pulmonary functions (Khaliq and Tripathi, 2009).

Omega-3 fatty acids, both plant-derived (α-linolenic acid, ALA) and forms found

primarily in fish (eicosapentaenoic acid, EPA and docosahexaenoic acid, DHA) (Deckelbaum and Torrejon, 2012) give rise to anti-inflammatory, pro-resolving mediators such as protectins, resolvins, and maresins and protect against pro-inflammatory stimuli (Duvall and Levy, 2016). Previous studies linked between higher dietary intake of omega-3 fatty acids decreases in cardiovascular risk/atherosclerosis (Jain et al., 2015), and the morbidity in asthma (Papamichael et al., 2018).

The aim of the work was to study the different effects of formalin inhalation on the structure of the lung and heart tissues and the possible protective role of omega-3 in experimental rats by histological and immune-histochemical studies.

**MATERIALS AND METHODS**

**Drugs:**
*Formaldehyde (FA)* 10% concentration solution was brought from El Nasr pharmaceutical chemicals Company, Egypt.
*Omega-3* is available in the form of liquid syrup; Sigma pharmaceutical industries, (Efalex 120 ml) composition per 5 ml: high DHA Fish oil 640 mg, Rigel evening primrose oil 213 mg, DL-alpha tocopherol acetate, thyme oil 0.40 mg, equivalent to vitamin E 7.82 mg.

**Animals:**
Thirty adult male albino rats, aging 4 - 6 months and weighing 200 - 250 gm. The animals were obtained from the Animal House, Faculty of Medicine, Assiut University, and were housed in the Animal Facility at Faculty of Medicine, Sohag University, Egypt. All rats were given access to a rodent chow diet and water. The experiment was performed according to the "Guide for the Care and Use of Laboratory Animals" Institute of Laboratory Animal Resources (2011) and in accordance with the guidelines of the University Animal Ethics and approved by the Research Ethics Committee considering care and use of laboratory animals.

**Experimental Design:**
After a 7-day of acclimatization, rats were equally divided into three groups as follows:

**Group I (Control Group):** It was composed of 10 adult male albino rats. They were not subjected to any treatment.

**Group II (Formaldehyde Exposure Group):** It included 10 adult male rats subjected to formaldehyde inhalation released from a cotton piece placed in a small glass box inside the cages and soaked with 10% FA solution. The cotton piece was replaced every one hour to keep a constant concentration. These animals were subjected to formaldehyde inhalation for 8 hours/day. This was done for 6 days/week for four weeks. (Sayed et al., 2013; Isaac and Saad, 2018).

**Group III (Formaldehyde and Omega-3 group):** included 10 adult male rats subjected to formaldehyde inhalation as Group II and orally administered with 300 mg/kg/day of Omega-3 for 4 weeks using sterilized rats stomach tube. (El Desouky et al., 2019).

After 24 hours from the last dose, rats were anesthetized using ether inhalation, carefully dissected, and the specimens from the lung and the left ventricle of the heart were taken.

**Preparation of The Specimens for Light Microscopic Examination:**
Perfusion fixation was used; the specimens were fixed in 10% neutral buffered formalin and processed for light microscopic study. Paraffin sections of 6μm thickness were obtained for Haematoxylin, and eosin, Masson trichome, and Immunohistochemical stains (Drury and Wallington, 1980).

**Immunohistochemical Methods:**
Immunohistochemical staining was carried by avidin biotin peroxidase complex method. The specimens from the lungs and cardiac muscles were processed. Caspase-3,
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(mouse monoclonal antibody and anticleaved caspase-3 rabbit polyclonal antibody (Neo Markers Fermont, CA 94539, USA) were used for detection of apoptosis. The reaction appeared brownish either cytoplasmic or nuclear. Sections were then counterstained with Mayer’s hematoxylin (HX), dehydrated, cleared, and mounted. Tonsils were used as positive control tissues. Negative control was performed after omitting the primary antibody (Remick et al., 2006).

Paraffin sections were also stained with avidin-biotin-peroxidase for demonstration of cells immunoreactive to TNF-α (Minneapolis, Minnesota, USA), and counterstained with HX. TNF-α expression was a cytoplasmic immunopositive reaction (Dabbs, 2002).

Morphometric Study and Statistical Analysis:
Collagen quantificationSemi-automated image analysis was applied (Schipke et al., 2017). From each Masson's trichrome stained section, five random fields for each group were selected and imaged using an objective lens magnification of 10x for lung sections, 20x for heart sections. ImageJ software (version 1.51k, Wayne Rasband, National Institutes of Health, USA) was used for the analysis.

Variables were represented by mean ± SD (Mean ± standard deviation of mean). The SPSS program version 16 was used to analyze the differences among all groups in all the data parameters by one-way analysis of variance and a post-hoc test was used to find the statistical difference between the groups when ANOVA was statistically significant (P value ≤0.05) (Dean et al., 2000).

RESULTS

Lung Tissue:

Anatomical Results:
The morphological appearance of the lungs of the control group (I) revealed the normal rosy pink color of the examined lobes with regular borders and normal shape. There was no change in size with no air cystic changes or abnormal masses (Fig. A).

The gross morphological changes in the lungs of group II (FA exposed) revealed congestion in all lobes with increased pulmonary vascular marking. The lungs appeared smaller in size with a prominent mass in one of the treated lungs (Fig. B). The gross morphological changes in the lungs of the group III (FA and omega-3 exposed) animals revealed mild congestion in some lobes with no significant changes in the size. The lungs had normal shape and borders with no air cystic changes or abnormal masses (Fig. C).

Histological Results:
Control group (I) revealed normal pulmonary tissue architecture with clear patent bronchial passages and alveolar cavities including the alveolar sacs, the alveolar ducts, and the alveoli. The alveolar septa had normal thickness with no abnormality in alveolar septal blood capillaries. Normal collagen fibers distribution in lung tissues (Figs. 1 & 4).

Group II (four weeks FA exposure) specimens confirmed the gross morphological changes with loss of normal lung architecture and destructed alveoli. There were focal pneumonia granulomas with massive inflammatory infiltrate and apparent hemorrhage in-between alveoli. Massive collagen distribution was found in lung tissues (Figs. 2 & 5).

Group III (FA and omega 3 exposed) showed restoration of normal lung architecture with normal appearance of alveoli and sinuses, Still, inflammatory infiltrate appeared mainly around blood vessels. Minimal collagen deposition appeared in the lung interstitium (Figs. 3 & 6).

Immunohistochemical Reaction:
For TNF-α: a negative reaction in the control group (Fig. 7) whereas in the group II positive immunohistochemical staining of TNF-α observed in alveolar interseptal region,
muscle cells surrounding the bronchi and bronchioles, and the epithelium of the bronchi and bronchioles (Fig. 8) Minimal positive immunohistochemical staining of TNF-α was observed in the alveolar interseptal region in the group III (Fig. 9).

For caspase-3: the control group, showed a mild positive reaction (Fig. 10). In the FA-exposed group, there was a massively positive reaction compared to the control group (Fig. 11). Whereas, FA and omega-3 showed moderate positive reaction (Fig. 12).

**Cardiac Muscle:**

**Anatomical Results:**

No gross abnormalities or changes observed between hearts in the three groups

**Histological Results:**

**Control group (I)** Light microscope showed cardiac myocytes in a longitudinal section with prominent striations. Cardiac muscle fibers had acidophilic cytoplasm and central, vesicular, and oval nuclei with a rich capillary network (Fig. 13), with scanty collagen fiber (Fig. 16).

**Group II (four weeks FA exposure)** showed irregular, shrunken, and widely separated cardiomyocytes with pyknotic nuclei. Hemorrhage and Inflammatory cells infiltration were observed in the widened interstitium (Fig. 14). Abundant collagen fibers were seen in-between the cardiomyocytes (Fig. 17).

**Group III (FA and omega 3 exposed)** showed restoration of the normal picture of the cardiac muscle with acidophilic cytoplasm and vesicular, oval nuclei (Fig. 15). A considerable amount of collagen fibers were seen in-between the cardiomyocytes (Fig. 18).

**Immunohistochemical Reaction:**

For TNF-α, a negative reaction in the control group (Fig.19), whereas in group II positive immunohistochemical staining of TNF-α was observed in-between cardiac muscle fibers. (Fig. 20) and negative immunohistochemical reaction of TNF-α was observed in group III (Fig. 21).

For caspase-3, the control group showed a negative reaction (Fig. 22). In the FA-exposed group; there was a positive reaction compared to the control group (Fig. 23). FA and omega 3 showed a negative reaction (Fig. 24).
A: Gross picture of lung of control rats showing normal rosy pink color of the examined lobes.

B: Gross picture of lung of FD treated rats showing marked congestion (stars), with prominent mass in one of the treated lungs (arrow).

C: Gross picture of lung of FD and Omega treated rats showing mild congestion (stars), with normal shape and borders of lungs.
Fig. 1: Transverse section of the lung of Control lung (group I) showing normal lung architecture with alveoli (A), alveolar sacs (S), thin (thin arrow) and thick (thick arrow) portions of the interalveolar septa, bronchioles (B), and blood vessels (BV). H&E X 100.

Fig. 2: Transverse section of the lung of group II (treated with formalin only) showing loss of lung architecture and destruction of alveoli (stars), dilated sinuses (S), infiltration of inflammatory cells (I) around the blood vessels, and hemorrhage (HG). H&E X 100.
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Fig. 3: Transverse section of the lung of group III (treated with Formalin and omega-3) showing restoration of normal lung architecture with normal appearance of alveoli (A) and sinuses (S). Still Inflammatory infiltrate (I) appears mainly around blood vessels (BV). H&E X 100.

Fig. 4: Transverse section of the lung of Control lung (group I) demonstrating normal collagen fibers distribution in lung tissues (arrows). Masson’s trichrome X 100.
Fig. 5: Transverse section of the lung of group II (treated with Formalin only) demonstrating extensive collagen deposition are recognized in the lung interstitium (arrows) and around bronchioles (stars). Masson’s trichrome x 100

Fig. 6: Transverse section of the lung of group III (treated with Formalin and omega-3) demonstrating minimal collagen deposition in the lung interstitium (arrow) and around bronchioles (stars). Masson’s trichrome x 100
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Fig. 7: Transverse section of the lung of Control lung (group I) showing negative reaction for TNF-α between lung tissues. TNF-α × 100

Fig. 8: Transverse section of the lung of group II (treated with Formalin only) showing a positive immunohistochemical staining of TNF-α observed in alveolar interseptal region (arrows), muscle cells surrounding the bronchi and bronchioles (irregular arrow), the epithelium of the bronchi and bronchioles (stars) NF-α (↑). TNF-α × 100
Fig. 9: Transverse section of the lung of group III (treated with Formalin and omega-3) showing minimal positive immunohistochemical staining of TNF-α observed in alveolar interseptal region (arrows). TNF-α × 100

Fig. 10: Transverse section of the lung of Control lung (group I) showing mild positive reaction for caspase-3 in the wall of bronchi and alveoli. Caspase-3× 100
Fig. 11: Transverse section of the lung of group II (treated with Formalin only) showing Massive caspase-3-positive cells were observed at the walls of bronchi and alveoli. Caspase-3× 100

Fig. 12: Transverse section of the lung of group III (treated with Formalin and omega-3) showing moderate reaction of caspase-3 observed at the walls of bronchi and alveoli. Caspase-3× 100
Fig. 13: Control heart (group I) showing: longitudinally arranged cardiac muscle fibers which have acidophilic cytoplasm and central, vesicular and oval nuclei (short arrow). They have obvious longitudinal striation cardiac fiber (CF). Note: Blood vessels (BV) are observed. H&E X 200.

Fig. 14: Formalin treated heart (group II) showing shrinkage and irregularity of cardiac fibers (star) which have acidophilic cytoplasm and pyknotic nuclei (short arrow) with hemorrhage in-between cells. H&E X 200.
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Fig. 15: Formalin and omega-3 treated heart (group III) showing longitudinally arranged cardiac muscle fibers which have acidophilic cytoplasm and vesicular, oval nuclei (short arrow) with obvious longitudinal striation cardiac fiber (CF). Also with normal appearance of Blood vessels (BV). H&E X 200.

Fig. 16: Control heart (group I) showing Scanty content of collagen fibers can be seen in-between the cardiomyocytes (↑). Masson’s trichrome x 200.
Fig. 17: Formalin treated heart (group II) **showing abundant collagen fibers seen in-between the cardiomyocytes** (↑) Masson's trichrome x 200.

Fig. 18: Formalin and omega-3 treated heart (group III) **showing a considerable amount of collagen fibers are seen in-between the cardiomyocytes** (↑). **Masson's trichrome x 200.**
Fig. 19: Control heart (group I) showing negative reaction for TNF-α in-between cardiac muscle fibers (section of cardiac muscles). TNF-α × 200.

Fig. 20: Formalin treated heart (group II) showing positive immune reaction for TNF-α (†) (section of cardiac muscles). TNF-α × 200.
Fig. 21: Formalin and omega-3 treated heart (group III) showing negative reaction for TNF-α between cardiac muscle fibers (section of cardiac muscles). TNF-α × 200.

Fig. 22: Control heart (group I) showing negative reaction for caspase-3 between cardiac muscle fibers (transverse section of cardiac muscles). caspase-3 × 400.
Fig. 23: Formalin treated heart (group II) showing positive immune reaction for caspse-3(↑). (transverse section of cardiac muscles) × 400.

Fig. 24: Formalin and omega-3 treated heart (group III) showing negative reaction for caspse-3between cardiac muscle fibers (transverse section) (transverse section of cardiac muscles). × 400
Morphometric Results:

Lung: The mean area percentage occupied by collagen in the formaldehyde group (24±4.8) was a very highly significant increase in comparison to the control group (14.2±3) \( p < 0.001 \). The mean area percentage occupied by collagen in the formaldehyde +omega group (17.7±3.27) was non-significant changed in comparison to control group \( p \leq 0.1 \) (Table 1) (Histogram 1).

Heart: The mean area percentage occupied by collagen in the formaldehyde group (24±2.5) was a highly significant increase in comparison to the control group (10.2±.4) \( p \leq 0.02 \). The mean area percentage occupied by collagen in the formaldehyde +omega group (20.9±3.5) was a very highly significant increase in comparison with control \( p \leq 0.007 \). (Table 1) (Histogram 1).

Table 1: showing the mean ± SD of area percentage of collagen fibers between lung tissue and cardiac myocytes in different groups (P ≤0.01 (**) → High significant difference, P≤ 0.001 (***) → Very high significant difference. ns→ Non significant).

<table>
<thead>
<tr>
<th>Groups</th>
<th>Control</th>
<th>Formaldehyde</th>
<th>Omega</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lung</td>
<td>14.2±3</td>
<td>24±4.8***</td>
<td>17.7±3.27ns</td>
</tr>
<tr>
<td>Heart</td>
<td>10.2±.46</td>
<td>24±2.5**</td>
<td>20.9±3.5***</td>
</tr>
</tbody>
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Histogram 1: Comparison between the studied groups according to the percentage of the area of collagen in lung and heart tissues

DISCUSSION

The study of the relationship between atmospheric pollution and respiratory health needs to take into account the anatomical, histological, and toxicological effects. Inhalation is the main exposure pathway of these pollutants, which makes the respiratory tract the first target organ of airborne pollutants (Takahashi et al., 2007; Bernstein et al., 2008).

In the present study, the gross morphological study of the lungs exposed to formaldehyde revealed irregular convoluted surfaces of most lungs, increased congestion, and appearance of masses. These results are in agreement with previous studies whereas rats were exposed to formaldehyde for different durations which showed congestion in most lobes and focal pneumonic organization (Khaliq and Tripathi, 2009; Afrin et al., 2016; El Desouky...
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et al., 2019).

Lung sections from the rats exposed to formaldehyde revealed pulmonary changes in the architecture of the lung and an increase in collagen contents, these changes were consistent with the findings in the rabbit lungs after exposure to 40% formaldehyde (Neelam et al., 2011), also detected in rats (Kamata et al., 1996; Turkoglu et al., 2008; Mehdi et al., 2014) and in mice (YU et al., 2004).

The positive reaction to TNF-α in the present study was confirmed by the previous results which showed that FA exposure mice enhanced TNF-α in the lung and increased the collagen production (Leal et al., 2018)

Increase reaction to caspase-3 in formalin exposed rats in the present study was confirmed by the previous results which improved that inhalation of formaldehyde induces apoptotic cell death in the lung tissues (Sandikci et al., 2009)

Cardiac sections from rats exposed to formaldehyde revealed changes in the architecture of the cardiac muscle and an increase in the collagen contents which was confirmed by the previous results (Afifi and Hanon, 2011) which revealed that rats exposed to oral formalin consumption showed disruption and fragmentation of cardiac myocytes and increased collagen fiber content between the separated cardiac muscle fibers. (Onyeka et al., 2018) showed that Formaldehyde fumes have cardiopulmonary effect on medical students. Formaldehyde caused cardiac failure possibly mediated by impairments of the intracellular Ca2+ handling in rat normal and hypertrophic hearts (Takeshita et al., 2009).

The positive reaction to TNF-α in the cardiac sections in the present study was confirmed by the previous results which proved that TNF-α is among the most common pro-inflammatory cytokines observed following cardiac stress (Gullestad et al., 2012; Bartekova et al., 2018).

The positive reaction to caspase-3 in the cardiac sections in formalin exposed rats in the present study was confirmed by the previous results (Yue et al., 1998) which showed that caspase expression and activation were reported in apoptosis of isolated rat cardiomyocytes. Guttenplan et al., 2001 found that caspases can act as effectors, participating in the total disassembly of cell structures and represents the principal form of myocyte death. Caspase-3 activity was proved to be responsible for the cardiomyocyte apoptosis in heart failure rabbit obtained by ventricular pacing (Laugwitz et al., 2001).

The addition of omega-3 in the present study revealed that restoration of the normal lung architecture with the normal appearance of alveoli and sinuses with minimal collagen deposition. These results were confirmed by the previous studies which showed that elevating tissue omega-3 levels can prevent and treat fine particle-induced health problems and suppress fine particle-induced pulmonary and systemic inflammation (Li et al., 2017). The therapeutic role of omega-3 was confirmed also by (Raafat et al., 2018) who found that the addition of omega-3 has an ameliorating effect on the inflammation caused due to rheumatoid arthritis on the lung tissue by H&E and (TNF- α). The anti-inflammatory effect of omega-3 was studied also by (xu et al., 2011) who said that omega-3 injection effectively reduces destruction and injury in the lung of rats with severe scald.

Omega-3 FAs are pivotal not only for brain function and normal growth and development (Belluzzi, 2002) but also they have many beneficial effects on several health problems including cardiac diseases (Roy and Le Guennec, 2017; Endo and Arita, 2016) which confirm the present results of restoring the cardiac muscle architecture after addition of omega-3.

Omega-3 FAs administration was also proved to prevent pathological alterations in the myocardium from Bisphenol exposure (Bahey et al., 2019).

Conclusion:

Formaldehyde inhalation has a sever destructive effect on both lung and heart tissues, the addition of omega-3 has an ameliorating effect on these effects.
REFERENCES


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ARABIC SUMMARY

تأثير الفورمالدهايد على بنية الرئة والقلب والتأثير الوقائي المحتمل لأوميجا 3 (دراسة هستولوجية ومستحيلة مناعية)

سنوي محمد عويس، عبير فريد عبد النعيم
قسم التشريح وعلم الأجنة، كلية الطب، جامعة سوهاج

مقدمة: الفومالدهايد هو ملوث داخلي وخارجي شائع يوجد في العديد من المنتجات. إن سمية الفورمالدهايد تثير القلق لجميع الذين يعملون بشكل وثيق معها مثل التحنيط، التشريح، فنيي علم الأنسجة وطلاب الطب. أوميجا 3 في زيت السمك هو واحد من أهم الأحماض الدهنية غير المشبعة التي تنشئ مضادات للأكسدة والالتهابات.

الغرض من البحث: دراسة تأثير استنشاق الفورمالدهايد على نسيج الرئة والقلب والتاثير الواقى المحتمل لأوميجا 3 المواد والأسباب: تم تقسيم ثلاثين من ذكور الفئران البيض إلى ثلاث مجموعات متساوية. المجموعة الأولى (المجموعة الضابطة)، تعرضت الفئران المجموعة الثانية لاستنشاق الفورمالدهايد لمدة 4 أسابيع. المجموعة الثالثة: تعرضوا لاستنشاق الفورمالدهايد كالمجموعة سابقة وأعطوا عن طريق الفم أوميجا 3 بجرعة 300 مكغ / كغ / يومًا لمدة 4 أسابيع. بعد 24 ساعة من الجرعة الأخيرة، تم تشريح الحيوانات. تم معالجة القلب والرئتين من صباغتها بالهيماتكسول والماسون ترايكوم والدراسة الهستولوجية المناعية.

النتائج: أظهرت الفئران المعالجة ب استنشاق الفورمالدهايد تغيرات مهمة في بنية كل من الرئة والقلب، وزادت نسبة ألياف الكولاجين الوكولاجين، وزاد التعبير المناعي لكل من caspase-3 و TNF-α. هذه الدراسة تختصر هذه التغيرات السريعة وظيفية وتأتي النتيجة الديناميكية للفورمالدهايد والتعبير المناعي لكل من الرئة والقلب.

الخلاصة: استنشاق الفورمالدهايد يرتبط بتغيرات ميكروسكوبية لكل من الرئة والقلب، إضافة أوميجا 3 يمكن أن تقلل من هذه التغيرات.